

Out of the darkness and into the light — Ushering in a new era of anti-inflammatory based medicine by maximizing the D6D metabolic pathway — Essential to reversing CVD & Cancer

Brian Scott Peskin *

- Affiliation; Peskin pharmaceuticals www.peskinpharma.com
- Correspondence: prof-peskin@peskinpharma.com

*Author to whom any correspondence should be addressed.

Abstract

This review article examines lipid physiology and its key role in advancing anti-inflammatory medicine; in particular, reversing CVD (the world's #1 killer). Despite recent media claims that omega-6 seed oils are harmful, adequate consumption of these lipids is essential for maximizing systemic anti-inflammation functionality and decreasing CVD; in particular, maximizing production of the body's most powerful anti-inflammatory, PGE₁. The issue lies exclusively in the harmful food / lipids processing.

Omega-6 is an Essential Fatty Acid (EFA). The body can't make it. In fact, both series of EFAs (omega-6 series and omega-3 series) *must* come from food. As the science makes crystal clear, lipids are the "brick and mortar" of our 100 trillion cells. They are required to optimize cellular membrane structure and cellular terrain, increasing cellular oxygenation, and decreasing today's epidemic of chronic inflammation. Parent omega-6 (LA) is the main EFA-based component of our 100 trillion bi-lipid cellular membranes (there is at least 10-fold more Parent omega-6 than Parent omega-3 throughout the tissues and organs). In the body, LA quantities far exceed ALA quantities or long-chain metabolites like EPA / DHA. Of prime importance, Prostaglandin Series 1 (PGE₁) — being the body's most powerful cellular anti-inflammatory — is derived from Parent Omega-6. Nothing in the omega-3 series matches PGE₁'s anti-inflammation effectiveness, increased vasodilation, or cellular oxygenating power. Chronic inflammation is now widely recognized as a fundamental element of the leading causes of death, including both cardiovascular disease and cancer. Additionally, chronic epidemics of numerous inflammatory conditions are associated with impairment of the delta-6 desaturase metabolic pathway (D6D). This dysfunctional impairment results in reduced PGE₁ production and chronic (persistent) systemic inflammation. Unless a new era of anti-inflammation-based medicine takes center stage, it is impossible to reverse these epidemics of disease.

Many pivotal lipid physiology research papers remain underpublicized. Without knowledge of and comprehension of these seminal works, it is impossible to fully understand intricate lipids-based connections. This review article provides novel insights into the importance of Parent omega-6 and newly available methods to mitigate D6D pathway impairment and maximize PGE₁ output. Emphasis on anti-inflammatory medicine is vital to reversing the escalating prevalence of both established and newly emerging diseases. Cardiovascular disease — the leading cause of mortality worldwide, and newly created inflammatory-based epidemics, such as type II diabetes and autism, exemplify this need. Notably, type II diabetes was virtually non-existent pre-1940, is now an epidemic, and autism, rare just a

few decades ago, is now also an epidemic affecting over 3% of all children. Both are now known to be inflammatory-based.

Fish/marine oil supplements have become extremely popular. However, when consumed in the quantities often suggested by healthcare professionals and then used by patients, the (supraphysiological) amounts of EPA/DHA consumed directly lead to significant chronic inflammatory issues. Marine oils alter the levels of LA in mitochondria and all other tissues, causing horrific results. By highlighting under-publicized lipids-based research, it is now possible to impartially and objectively scientifically address and reverse the rising prevalence of chronic inflammatory diseases. This article uses cardiovascular disease as the prime clinical example, but the same concepts apply to mitigate the severity of the numerous diseases now known to have a significant, even causal, inflammatory component.

Keywords: essential fatty acids; delta-6 desaturase; eicosanoids; parent essential fatty acids; parent essential oils; PEOs; PUFA; polyunsaturated fatty acids

1. Introduction: Key Biological Lipids General Background — Parent EFAs (PEOs) and Derivatives

Linoleic acid (LA) and alpha-linolenic acid (ALA) are the only true 18-carbon chain essential fatty acids. They cannot be converted into each other nor synthesized by the body and must be obtained from dietary sources. LA is "Parent" omega-6, and ALA is "Parent" omega-3. Metabolites (eicosanoids), derived from LA and ALA, are longer-chain structures produced by the body, as needed, but are not essential because they can be synthesized by the body, and therefore, NOT EFAs. They are properly termed EFA derivatives. Eicosapentaenoic acid (EPA) and docosahexaenoic acid (DHA) are derivatives. In this review, I refer to LA and ALA as "Parent Essential Oils" (PEOs) or "Parents," while longer-chain metabolites (and eicosanoids) are termed "derivatives." The body produces these derivatives from Parents in small, minuscule amounts, as needed. Literature often confuses these two distinct categories, even inappropriately "lumping together" both Parents and Derivatives.

Cellular membrane lipid physiology

Each of a human's 100 trillion cells consists of a bi-lipid membrane. Importantly, the essential PEOs comprise 25% - 33% of their polyunsaturated membrane lipids.ⁱ Additionally, every mitochondrion, typically hundreds to

thousands per cell, contains PEOs; in particular, Parent omega-6.^{ii,iii} Therefore, PEOs must be considered the "brick and mortar" of every cell, tissue, and organ, including mitochondria. In sharp contrast, aside from the brain, eyes, and nervous system, most tissues and organs contain few derivatives like EPA/DHA in their cellular membranes. By far, the most significant method to physiologically change tissue structure / physiology is through its lipid structure.^{i,iii, iii}

2. Organic / Fully Functional / Unprocessed vs Adulterated / Processed EFAs

Not distinguishing an adulterated (processed) nonfunctional EFA-containing lipid versus a fully functional *unprocessed* fully functional EFA-containing lipid—in particular, omega-6 series LA—is the *prime cause* of confusion leading to both inconsistent clinical trials and poor patient outcomes. The criticality of distinguishing between the effects of adulterated vs unadulterated forms of LA is obvious. Failure to feed rodent chow (and humans) with fully functional LA food has led to the incorrect and misleading conclusion that dietary intake of LA increases disease risk, including CVD risk.^{iv} Decades ago, we personally verified this adulteration of mouse chow in a cancer study. We had a sample of mouse chow measured for oxidation. The rancidity level was at least 3Xs greater than the lowest allowable safe rancidity peroxide value (PV) measurement level, i.e., the mouse chow itself was cancer- / heart

disease-causing. The researchers routinely have no idea of this physiologic lipid distortion skewing their trials.

With functional LA deficiency — caused by consuming processed omega-6 cooking oils — there is an enormous increase in permeability of the skin (epithelial tissue) and an increase in capillary fragility, further explaining the pathophysiology of CVD and various skin disorders, along with the path to prevention.^v Oxidation of LDL-C causes significant depletion of functional cellular LA (Parent omega-6).^{vi} This oxidation occurs *ex vivo* from processed oils used in cooking.

By not understanding this lipid physiology, patients are *unknowingly* harming themselves. For example, by ingesting fish oil (EPA/DHA), there is a corresponding decrease in tissue's quantity of LA, causing significant pathophysiologic deficiency of fully functional Parent omega-6 in tissues / organs.^{vii}

Of particular note, with ingestion of fish oil (EPA/DHA), there was a corresponding decrease in tissue's LA, adding to pathophysiologic levels of cellular deficiency.^{vii}

3. PEO Quantities—Omega-6 Dominates

With the extensive focus on omega-3 series fatty acids today, like EPA / DHA, it is significant to note that the free Parent fatty acids (non-esterified) in human plasma, although minute in quantity, are ordinarily composed of about 15% LA (linoleic acid, Parent omega- 6) and a miniscule 1% of ALA (alpha linolenic acid, Parent omega-3).^{viii} Derivatives such as EPA/DHA are naturally significantly less in quantity than LA. In sharp contrast to the high amounts of n-6 series PUFAs, n-3 series PUFAs account for only 1.8% of the fatty acids in triglycerides, 3.5% in the phospholipids, and only 1.7% (ALA is 0.5%) in cholesterol esters. However, the Parent (LA/ALA) ratios in triglycerides are 23:1. N-3 PUFA makes up only 1% - 2% of fatty acids in plasma.^{ix} Even in the brain, the LA / ALA uptake (ratio) is an amazingly 100 times greater in favor of LA.^x

4. Variable Tissue Composition

The significant variable in tissue is its lipid structure. Although the genetics of a particular species precisely specify cellular structure, its lipid composition can vary significantly based on EFA-containing food consumption —in particular, when supra-pharmacologic amounts of long-chain metabolites are consumed — such as the case with fish/marine oil supplements. A pharmacologic overdose can't be oxidized away for energy or otherwise. Consequently, much of this overdose is forced into tissue composition, such as cardiolipin in the mitochondria and the skin, causing an improper tissue composition, often maintaining a linear relationship of the overdose in the plasma, liver, and RBCs.^{xi,xii,xiii} This physiological fact is also underpublicized. Cellular bi-lipid membrane structure and the LDL cholesterol structure warrant intense investigation.

Lipid Physiological Variability in LDL-C

The structure of LDL-C is complex. Its cholesteryl ester is key (**Figure 1**). The structure of cholesterol itself never changes. What changes is exclusively its *esterified* moiety—the acyl side chain. That's a critical difference that many in the medical community may not appreciate. This is a simple condensation reaction, removing the water, catalyzed by the enzyme ACAT (Acyl CoA: Cholesterol Acyl Transferase) between a fatty acid and cholesterol. "R" symbolizes the hydrocarbon portion of the fatty acid. For example, if oleic acid were esterified with cholesterol, then R would be - C7H14CH=CH-C8H17 with the double bond in *cis* configuration.

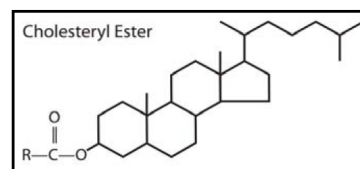


Figure 1. Cholesterol Ester. Lipoproteins transport cholesterol and its esterified PEOs to the tissues via apoprotein B-100 (ApoB100).

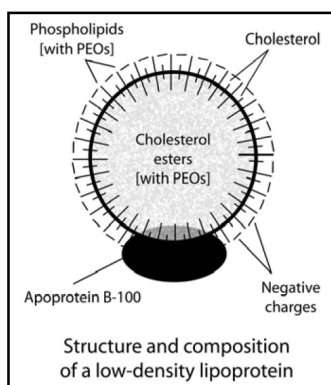


Figure 2. Structure and composition of a low-density lipoprotein showing the significance of its esterified cholesterol structure.

5. LDL-C Is NOT Oxidized in the Body / Bloodstream: It is Oxidized from Food Processing

Cholesterol itself is extremely resistant to oxidation, whereas its main esterified component, Parent omega-6 (LA), is much more easily oxidized, especially *ex vivo*. Dietary LA that has already become oxidized prior to ingestion *ex vivo* is ubiquitous through the processing of foods or overheating, since heating in the presence of air enhances peroxidation of PUFA glycerol esters.^{xiv,xv}

It is highly unlikely that LDL can become oxidized in plasma to the extent that it causes foam cell formation and possesses chemotactic and cytotoxic properties. Furthermore, only minimal physical and chemical changes related to oxidation are produced by even a prolonged storage of LDL with oxygen or by incubation with low concentrations of copper ions. Clearly, the quantity of antioxidants is too small for oxidation *in vivo* to be a significant physiologic issue.^{vi} Nature doesn't require high quantities of antioxidants in this area because the substances are naturally resistant to oxidation in the body. The PUFA, in particular, LA, is being consumed and entering the body in a dangerous oxidized state.

Confirmation of Exogenous Damage to Parent Omega-6 Prior to Consumption

This lipidology is confirmed by Prof. Gerhard Spiteller, former Chairholder of Biochemistry, Institute of Organic Chemistry at the University of Bayreuth, Germany. He has extensively

investigated EFAs and their degradation products, specifically, the influence of these substances on the physiology of mammals. He concluded that consumption of oxidized PUFA-cholesterol esters (from food processing) is responsible for the initial damage to endothelial cells, leading to premature cardiovascular disease. Cholesterol oxidation products are incorporated into LDL cholesterol in the liver.^{xvi} Ultimately, tissue injury is not caused by an increase in free cholesterol but by an increase in the cholesterol esters — the PEOs “magnetized” to the cholesteryl molecule for transportation in the bloodstream.^{xvii}

As a clinical example, in atherosclerotic patients, LDL cholesterol is altered /adulterated *ex vivo* by oxidative food-processing, and this altered LDL is taken up in unlimited amounts by macrophages. Dead macrophages filled with cholesterol's previously damaged, functionally impaired esters are then deposited in arteries. LDL-C is effectively transmitting a poison, i.e., nonfunctional and harmful LA. We can now explain the failure of statins to eliminate cardiovascular disease and stop CVD as the world's #1 killer.

By statins lowering of LDL-C, their esterified PEOs are also lowered, both adulterated [good outcome] and fully functional [bad outcome]. This is problematic. By focusing on displacing the *ex vivo* LA that has already become oxidized prior to ingestion through the processing of foods, cooking, or overheating, a solution can be found to mitigate this damage. Consuming organic / unprocessed / fully functional LA overpowers the adulterated LA; the oxidation issue is solved.^{xiii}

6. Diseases, Disorders of the Δ-6 Desaturase Pathway Causing Chronic Inflammation

Known inflammatory-based diseases and disorders associated with an impaired Δ-6 desaturase pathway include: Diabetes (both Type 1 & Type II) including associated neuropathy; lipid enveloped viruses (including COVID series); dermatological conditions (Eczema, etc.); cardiovascular disease (including soft plaque, hard, calcified plaque, hypertension, etc.); inflammatory bowel disease; chronic fatigue; fatty liver disease, including NAFLD; Multiple Sclerosis; Dementia /

Alzheimer's; Cancer, and respiratory diseases like COPD.^{xviii}

Specific Diseases Newly Re-classified as Inflammatory-Based (a partial selection):

Dementia / Alzheimer's

Although underpublicized, in 2018, Dementia and Alzheimer's have been newly reclassified as directly caused by inflammation, and considered a cardiovascular disease:

- “Inflammation is a MAJOR CAUSE, not just a consequence.... but only now is it identified as THE CAUSE. The new work turns previous thinking around.”^{xix}

Cardiovascular Disease

Tragically, no field better than cardiology exemplifies *the incredible lack of understanding* of the physiologic effects of lipids. Functional lipids are the key to decreasing cardiovascular disease. The lipid chromatography has been completed and published, if anyone cares to look. We know that LDL-C is the transporter of the esterified lipids. To reverse heart disease, there are two key lipidology-based points that have to be understood:^{xx,xxi,xxii}

- There is NO saturated fat in an arterial occlusion (clog).
- As much as 85% of the composition in an arterial occlusion is inflammatory, adulterated / processed Parent omega-6.

Calling omega-6 “inflammatory” is a mistake. This grave mistake came about because one of the components of omega-6 fatty acids, called arachidonic acid, is a (mistakenly termed) “building block” for various inflammation-related molecules. This has led to concern that omega-6 consumption would lead to a greater risk of heart disease. This has been completely disproven, and in 2009 it was stated by no less than the American Heart Association:^{xxiii}

- “*That reflects a rather naive understanding of the biochemistry,*” says

William S. Harris, Director of the Metabolism and Nutrition Research Center of the University of South Dakota Sanford School of Medicine and the nutritionist who led the science advisory committee that issued the report in *Circulation*.

- “[O]mega-6 PUFAs [Derivatives] also have powerful anti-inflammatory properties that counteract any proinflammatory activity,” say the advisory authors. ***It’s incorrect to view the omega-6 fatty acids as “proinflammatory.”***

Confirmation was proven by C-Reactive Protein marker (CRP), if anyone would care to look:^{xxiv}

- “Conclusions: Serum n-6 PUFAs [AA, etc.] were *not associated* with increased inflammation in men. In contrast, the main n-6 PUFA linoleic acid [Parent omega-6] had a **strong inverse association with the key inflammation marker CRP. “Omega-6 fatty acids do not promote low-grade inflammation.**
- “**The higher the serum linoleic acid [Parent omega-6] level, the lower the CRP. This is an inverse correlation.**”

How much more incorrect can the medical community be in their current recommendations concerning heart disease? A low-fat diet is still often recommended — a tragedy causing derailment off the path to inflammatory-based medicine.

Prostacyclin (PGI₂) is the body's most powerful natural vascular anticoagulant and is a powerful derivative of arachidonic acid (AA).^{xxv} Many, if not most, cardiologists I speak with are unaware of these older, yet undisputable and seminal findings. If these key lipidology facts aren't well-known, understood, and made use of, stopping the epidemic of the world's #1 killer can never occur.

Cancer

Much like the failure of CVD therapies to prevent the ever-increasing levels of heart attacks, the “war on cancer,” started in 1971 in the USA, has also been a dismal failure. Again, a woeful understanding of lipidology extends to this area, too. Cancer is not a genetic disease, and the coiner [Weinberg] of the term “oncogene” reversed his theory and now highlights inflammation as cancer’s cause back in 2007, if anyone would care to look.^{xxvi}

- Cancer researcher Robert Weinberg of MIT states: “The connection between **inflammation and cancer** has **moved to center stage** in the research arena.
- “...[I]**nflammation is the fuel** that feeds it [the malignant cancer].
- “In this **rewriting of the textbook**... This new view implies that rooting out every last cancer cell in the body might not be necessary. **Anti-inflammatory cancer therapy instead would prevent pre-malignant cells from turning** fully cancerous or would **impede an existing tumor from spreading** to distant sites in the body. Cancer victims might then be able to survive.”

Many oncologists — if not most oncologists, are unaware of this groundbreaking finding. Can this failure of the old guess and new anti-inflammatory-based path to eradicate cancer be any clearer? No. Without state-of-the-art lipidology, we will never “win the war on cancer,” regardless of how much money is “thrown” at defeating cancer. Furthermore, **ALL (100%) cancer tumors possess abnormal cardiolipin structure in their mitochondria**. EFA-wise, cardiolipin’s EFA structure is supposed to be comprised 100% of Parent omega-6 (LA), but without adequate, fully functional amounts, or excessive EPA / DHA from fish oil, cardiolipin’s structure will significantly be impaired.^{xxvii}

7. Fish Oil Impedes Cardiac Mitochondrial Functionality by Forcing Out Critically Required LA

Fish oil impairs mitochondrial cardiolipin functionality, as this seminal, yet underpublicized, *Journal of Biological Chemistry* finding makes clear:^{xxviii}

- “(18:2) CL₄ [Parent omega-6] rescues [fixes the damage] the major remodeling in the cardiolipin lipidome induced by long-term intake of DHA. [Cardiolipin is in the inner mitochondrial membrane.] Mitochondria are the cellular energy sources. Deficiency also causes chronic exhaustion – the #1 complaint of Americans.]
- “...[I]t is not the loss of linoleic acid alone that drives the impairment in enzyme function since the Western diet alone did not impair enzyme activities. Instead, it was the replacement of linoleic acid with DHA that promoted the reduction in activities.”

Could it be any clearer? No. Fish oil therapy is a horrific treatment for heart-related disease. Lowering of triglycerides by no means compensates for a heart with insufficient energy, leading to congestive heart failure.

8. Membrane Lipid Structure is Key to Decreased Cellular Inflammation

Another underpublicized yet what should have been an “*earth-shattering*” *journal article*, detailed how “[S]ecretory cells [virtually all cells] are *hypersensitive to their membrane lipids induced by the diet*.”^{xxix} This gives us the necessary link between *chronic, persistent inflammation* at the cell level and the skyrocketing inflammatory-based epidemics of disease. There is a thermodynamics-based reason that the cells can sense adulterated membrane lipids displacing the fully functional ones.

Today, there is almost an exclusive focus on proteins, with little attention on lipids. This seminal article makes clear that ***lipids control the proteins***. Lipids are the “protein masters.” Adulteration /

processing of cooking oil lipids precisely explains this epidemic increase in chronic cellular inflammation.

If there is a deficiency of fully functional LA in the diet, the body will substitute into cell membranes non-functional LA or even a nonessential fatty acid, such as oleic acid (non-essential omega-9), found in olive oil. This *forced* substitution because of inadequate functional LA results in a marked decrease of cellular oxygen transport with adverse effects on cellular metabolism and function.^{xxx} Because LDL cholesterol is the transport vehicle for PEO delivery into the cell, LDL cholesterol will transport any kind of LA into cells—defective or not—such as oxidized or *trans* entities (See Figure 1).

Adulterated / nonfunctional dietary LA, deposited in arterial intimal cell membranes, leads to abnormal oxidation at the vascular injury site, thus causing injurious inflammation. In this case, abnormal oxidation, caused by *ex vivo* adulteration of LA, involves formation of a hydroperoxide from LA by abstraction of a hydrogen atom as a radical from the doubly allylic methylene group between the two double bonds, followed by the addition of oxygen, a diradical, to make a hydroperoxide radical, which can then pick up another reactive hydrogen atom, perhaps from another LA molecule, to form the hydroperoxide. This, in turn, may break the O-O bond to form an alkoxide and a hydroxyl radical, which can continue to make more undesirable oxidized products []^{xxxi}. Therefore, atherosclerosis (the world's #1 killer) can be prevented / arrested if endothelial cells are fully functional by utilizing bioavailable Parent omega-6.^{xxxii}

9. Importance of the D6D Anti-Inflammatory Pathway

The delta-6 desaturase metabolic pathway is impaired in the majority, if not all, chronic inflammatory diseases.^{xxxiii} Once impaired, this desaturating pathway is not known to be reversible. Fortunately, there is no impairment in the elongase pathway; only in the desaturase pathway (Figure 3). We can help nutritionally compensate for this desaturase impairment by two methods: a) Increase

dietary fully functional, Parent Omega-6, and b) Bypass the D6D pathway increase by utilizing naturally occurring GLA-containing seed oils.

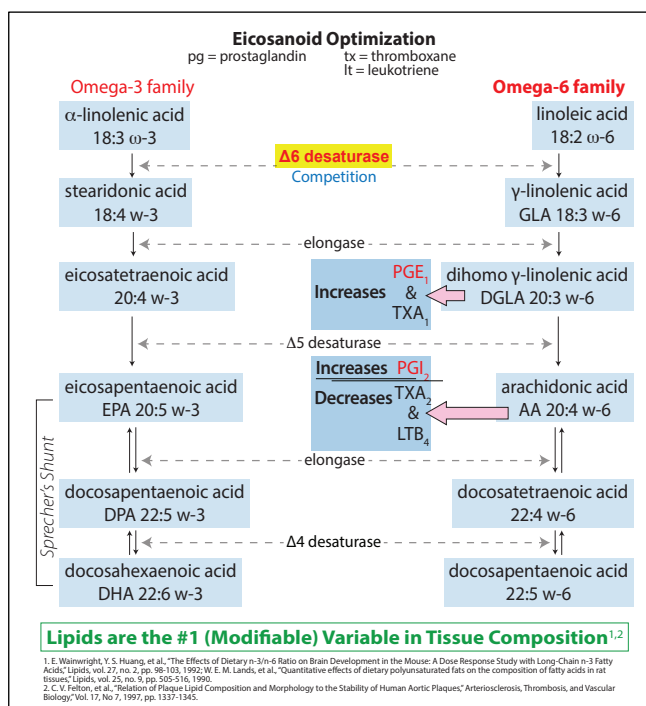


Figure 3. Parent Omega-6 and long chain metabolites (eicosanoids)

The majority of the plasma fatty acids are LA (Parent omega-6). Mitigating the damage caused by *ex vivo* intake of oxidized LA is now possible. Easiest is compensation by ingesting supplemental, fully functional, unadulterated, non-oxidized LA. Importantly, the key metabolites of LA—in particular, PGE₁ and PGI₂ (prostacyclin)—are significant vasodilators, increasing critical blood flow along with associated cellular oxygenation and nutrients. PGE₁ is also a potent anti-inflammatory and immune system regulator.^{xxxiii} If functional LA bioavailability is lowered, the potential for inflammation significantly rises, leading to atherosclerosis, etc.

Weiss, for example, has noted that PGE₁ (produced from functional Parent omega-6) reduces the fibrin deposition associated with the pathogenesis of atherosclerosis.^{xxxiv} Membrane fluidity increases when more functional (undamaged) polyunsaturated fatty acids—in particular, linoleic acid—are available to incorporate into the membrane lipid bilayer,

causing the epidemic of inflammatory-based diseases. Further underpublicized is that lipids are the #1 modifiable in tissue (*See below*).

No Delta-6 / -5 Desaturase Impairment in Healthy Patients Converting Parent Omega-3

Highly accurate, quantitative experiments were performed showing that the average healthy person and animals are both quite capable of metabolizing adequate amounts of DHA from Parent omega-3 (ALA). In a key NIH experiment, rodents naturally produced 50-fold (50Xs) more DHA each day than their brains required.^{xxxv} Certainly, Nature would ensure humans the same margin of safety shown to a rodent.

An *American Journal of Clinical Nutrition* article detailed over 60 firefighters and analyzed their conversion of omega-3 long-chain metabolites from Parent omega-3 (ALA), finding conversion adequate with sufficient intake of ALA [Parent omega-3].^{xxxvi}

Even vegans consuming no animal food, including fish, a group that absolutely would be expected to manifest gross neurological abnormalities, including both visual impairment and cognitive impairment, do not. There is no clinical evidence of such abnormalities in vegetarians.^{xxxvii,xxxviii} Confirmation in 2010 showed that vegetarians with an intake of just 0.3% DHA compared with fish eaters, produced 85% of the EPA levels and 83% of the DHA levels that consumers of fish did. These amounts are well within the “normal” ranges.^{xlii} There is no widespread impairment in the typical patient whatsoever; the normal conversion amounts are simply very low, naturally.

10. The Etiology of Cardiovascular Disease: Composition of Arterial Plaque

Current anti-CVD recommendations lack a firm physiologic / biochemical lipids basis. In 1994, using high-resolution chromatography, investigators found that plaque contained more than **10 different compounds, none of which were related to saturated fat.**^{xxiv,xxxix,xl} Not surprisingly, cholesterol was found in the plaque. This key finding demonstrated that cholesterol, esterified

with nonfunctional linoleic acid (LA)—adulterated Parent omega-6 — was by far the most abundant component in plaques of arterial stenosis. Furthermore, it was also found that cholesterol esters are the predominant lipid fraction in all plaque types, and that oxidized derivatives are toxic to most types of arterial cells.^{xlv}

Fish Oil Causes Decreased Prostacyclin Production Leading to Atherosclerosis

Prostaglandins are capable of both limiting thrombosis and reversing thrombosis in atherosclerotic patients.^{xli} Prostaglandin PGE₁ is the body’s most powerful anti-inflammatory and vasodilator, and prostacyclin (PGI₂) is a vasodilator that prevents both platelet adhesion and aggregation. These prostaglandins are both omega-6 metabolites. To the contrary, fish / marine oils increase endothelial platelet aggregation in atherosclerotic patients.^x In patients with atherosclerosis, prostacyclin (produced in endothelial tissue) biosynthesis fell by a mean of 42% during the fish-oil period, leading to increased adhesion against the arterial walls. [extremely bad outcome]. This finding is underpublicized. Synthesis of the platelet agonist thromboxane A₂ (produced in the platelets) declined by 58% [good outcome]. Atherosclerotic patients require increased intimal PGI₂ output, not decreased output.^{xliii} Furthermore, with marine oil, template bleeding times were significantly prolonged in all patients [bad outcome].

With Dietary Lipid / Eicosanoid Manipulation Lipid Physiology, Atherosclerosis Is Impeded Via Multiple Metabolic Pathways

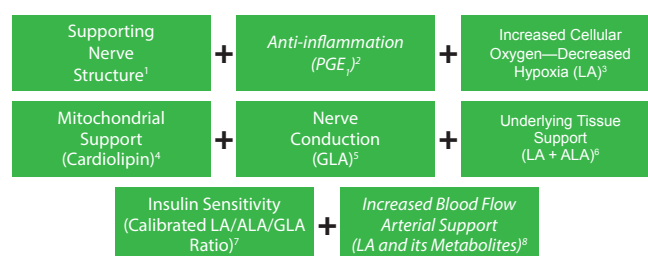


Figure 4. CAC Progression Impeded. Multiple Supporting Metabolic Pathways Optimized. References: i; xliii; xlvii; xlviii

11. Horrific Pathophysiology of Fish / Marine Oil Consumption in Humans

Fish oil spontaneously oxidizes at room temperature and in vivo

Lipid science clearly shows fish / marine oil is expected to contribute to CVD, not prevent it: a) Regardless of the anti-oxidant level added to the fish oil supplement, rancidity / peroxidation upon ingestion is a significant and problematic issue. Because of its high number of bis-allylic bonds — five double bonds in EPA and six double bonds in DHA, these metabolites are highly sensitive to temperature. Spontaneous oxidation of EPA leads to generation of a mixture of aldehydes, peroxides, and other oxidation products. Highly polyunsaturated, long-chained EPA and more so with DHA, due to its additional double-bond, is readily oxidized at room temperature (auto-oxidation) even in the absence of exogenous oxidizing reagents.^{xlix} Importantly, *in vivo*, a large increase in tissue and plasma accumulation of fatty acid oxidation products is noted in subjects consuming fish oil, even after the addition of antioxidant supplements to the diet. This deleterious effect led to a 14% decrease in life expectancy in those animals fed fish oil.¹ As shown above, PEOs don't suffer from this problematic *in vivo* oxidation issue. **In fact, DHA is 320Xs more susceptible to attack than common mono-unsaturated oleic acid (18:1) – like olive oil with no bis-allylic hydrogen bonds.** Fish / marine oil's EPA / causes excess oxidative stress.^{li} DHA Membrane lipid peroxidation should not be perceived solely as a 'damage to membranes' scenario but also as a significant endogenous source of damage to other cellular macromolecules, such as proteins and DNA (including mutations).^{”lii}

Another shocking, yet underpublicized, finding was that in primates and humans, such as the monkey, no quantity of *in vivo* antioxidants stops EPA/DHA damage as measured by lipofuscin, the peroxidized “age spots.” Lipofuscin was threefold (3Xs) greater in the livers of monkeys fed fish oil. Furthermore, another measure of oxidative damage, the basal thiobarbituric acid reactive substances (TBRS) levels, was four-fold (4Xs) greater than in

the monkeys fed (processed) corn oil with no EPA/DHA. The researchers found that even a ten-fold (10Xs) increase in alpha-tocopherol, a potent antioxidant, was not fully able to prevent the peroxidative damage from fish oil.^{liii} **It was known in 2000 that fish / marine oil, even in low doses, suppresses the innate immune system.**^{liv} Fortunately, not taking years, it takes 18 weeks to fully rid patients of fish oil in the cellular membrane.^{lv}

12. Parent Omega-6 to Parent Omega-3 Tissue Composition.

The preponderance of the omega-6 series throughout the body is made clear.^{lvi}

Table 1. Preponderance of Parent EFAs LA / ALA Ratios in Tissues & Organs.^{lix}

<i>Ratio of LA: ALA Tissue Composition</i>			
Tissue	% of Total Body Weight	Omega 6 PEO	Omega 3 PEO
Brain/Nervous System	3	1	1
Skin	4	1000	1
Organs & Other Tissues	9	4	1
Adipose Tissue (Body Fat)	15-35	22	1
Muscles	50	6.5	1

Parent-to-Derivative Metabolism and Amounts

What percentage of PEOs are converted to long-chain metabolites such as GLA, AA, EPA, DHA, etc.? This most important issue is rarely addressed. The USDA and NIH provide these answers. The conversion amount is much less than the medical field assumes; it is less than 5%—often less than 1%—with at least 95% of PEOs staying in Parent form. They are the “brick and mortar” of our 100 trillion cells. This singular mistake made decades ago in assuming very high conversion amounts, whereas in actuality they are extremely low conversion amounts, led to the irrational fish oil mania and wrong recommendations to limit Parent omega-6 cooking oil consumption; even the essential, fully functional / *unadulterated*.

Contrary to incorrect dogma, the enzymes that produce PEO derivatives (the delta-6 and delta-5 desaturase enzymes) are not significantly impaired in healthy patients.^{lvii} **Conversion of ALA [Parent**

omega-3] to DHA is unlikely to ever normally exceed a mere 1% with less than < 0.1% conversion to DHA.^{lviii} Research at the United States Department of Agriculture's USDA food composition laboratory (2001) reported a natural net conversion rate of a mere 0.046% of ALA to DHA & 0.2% to EPA—not the highly misleading often-quoted 15% conversion rate.^{lix}

It is important to understand that years ago, NIH researchers determined the amount of DHA utilized in human brain tissue to be **a mere 3.8 mg ± 1.7 mg/day**. Therefore, brain tissue in 95% of all subjects, allowing for variation in brain size, would consume merely 0.4 mg - 7.2 mg of DHA per day.^{lvii} New, twenty-first-century quantitative research from both NIH and USDA shows considerably lesser amounts of natural DHA conversion / usage from ALA than the medical community has been led to believe. These conversion amounts are extremely small and naturally limited. ***This dreadful mistake often leads to recommendations that are supra-pharmacologic and can potentially overdose patients by factors of 20-fold to 500-fold,*** depending on the specific supplement and amounts consumed. The body cannot simply oxidize these tremendous overdoses of EPA/DHA; they are too great a quantity. Their vast quantity displaces the required LA in tissue, as demonstrated in the congestive heart failure example,^{xxx} and the new epidemics of skin disorders like eczema in children in the US.

Amounts of Derivatives EPA / DHA in Fish Oil Supplements

An average 1000 mg health-food-grade fish oil capsule contains approximately 180 mg EPA and 120 mg DHA. Pharmaceutical-grade versions contain higher doses. The American Heart Association states that those with documented CHD are advised to consume about 1 gram of EPA + DHA per day. **Is this advice rational? No.**

ⁱ B. Alberts, D. Bray, J. Lewis, M. Raff, K. Roberts and J. Watson, "Molecular Biology of the Cell," 3rd Edition, Garland Science, New York, 1994, p. 428.

ⁱⁱ R. K. Murray, D. K. Granner, P. A. Mayes and P. A. Rodwell, "Harper's Illustrated Biochemistry," 26th Edition, McGraw-Hill, New York, 2003, p. 97.

Supplemental DHA from fish / marine oil not required

More underpublicized information (to reiterate):

- DHA is made by the body "as needed," from Parent omega-3 (ALA), although in extremely small quantities from Parent omega-3 (ALA). Highly accurate, quantitative experiments were performed showing that the average healthy person and animals are both quite capable of metabolizing adequate amounts of DHA from Parent omega-3 (ALA).
- In a key experiment, rodents naturally produced 50-fold (50Xs) more DHA each day than their brains required.^{xxxix} Certainly, nature would ensure humans the same margin of safety shown to a lowly rodent. Importantly.
- It is known that rodents metabolize EFAs the same as humans.^{xxxix}

Conclusion: By understanding and fully exploiting solutions to an impaired D6D metabolic pathway, a new era of anti-inflammatory medicine is now available. "Connecting the dots" of several, yet seminal, underpublicized lipid science discoveries gives us the precise answer to combat the current and increasing epidemics of chronic inflammatory-based diseases; in particular, CVD and cancer. This review article provides this underpublicized science.

Disclaimer/Publisher's Note: The statements, opinions, and data contained in all publications are solely those of the individual author(s) and contributor(s).

ⁱⁱⁱ C. Guyton and J. E. Hall, "Textbook of Medical Physiology," 9th Edition, W. B. Saunders Co., Philadelphia, 1996, pp. 16, 861-862.

^{iv} S. D. Anton, K. Heekin, C. Simkins and A. Acosta, "Differential Effects of Adulterated versus Unadulterated Forms of Linoleic Acid on Cardiovascular Health," Journal of

Integrative Medicine, Vol. 11, No. 1, 2013, pp. 2-10. doi:10.3736/jintegrmed2013002

^v H. M. Sinclair, "Deficiency of Essential Fatty Acids and Atherosclerosis, Etcetera," *Lancet*, Vol. 270, No. 6919, 1956, pp. 381-38.

^{vi} S. Young and S. Parthasarathy, "Why Are Low-Density Lipoproteins Atherogenic?" *Western Journal of Medicine*, Vol. 160, No. 2, 1994, pp. 1-18.

^{vii} M. B. Katan, J. P. Deslypere, A. P. van Birgelen, M. Penders and M. Zegwaard, "Kinetics of the Incorporation of Dietary Fatty Acids into Serum Cholesteryl Esters, Erythrocyte Membranes, and Adipose Tissue: An 18-Month Controlled Study," *Journal of Lipids Research*, Vol. 38, No. 10, 1997, pp. 2012-2022.

<http://www.ncbi.nlm.nih.gov/pubmed/9374124>

^{viii} H. Esterbauer, G. Jürgens, O. Quehenberger and E. Koller, "Autoxidation of Human Low-Density Lipoprotein: Loss of Polyunsaturated Fatty Acids and Vitamin E and Generation of Aldehydes," *Journal of Lipid Research*, Vol. 28, 1987, pp. 495-509. www.jlr.org/content/28/5/495.full.pdf

^{ix} A. A. Spector, "Plasma Free Fatty Acid and Lipoproteins as Sources of Polyunsaturated Fatty Acid for the Brain," *Journal of Molecular Neuroscience*, Vol. 16, No. 2-3, 2001, pp. 159-165.

^x H. Knapp, I. Reilly, P. Alessandrini and G. A. FitzGerald, "In Vivo Indexes of Platelet and Vascular Function during Fish-Oil Administration in Patients with Atherosclerosis," *The New England Journal of Medicine*, Vol. 314, No. 15, 1986, pp. 937-942. doi:10.1056/NEJM198604103141501

^{xi} P. E. Wainwright, Y. S. Huang, B. Bulman-Fleming, et al., "The Effects of Dietary n-3/n-6 Ratio on Brain Development in the Mouse: A Dose Response Study with Long-Chain n-3 Fatty Acids," *Lipids*, Vol. 27, No. 2, 1992, pp. 98-103. doi:10.1007/BF02535807

^{xii} S. K. Abbott, P. L. Else and A. J. Hulbert, "K. Abbott, P. L. Else and A. J. Hulbert, "Membrane Fatty Acid Composition of to the Balance of Dietary n-3 and Rat Skeletal Muscle Is Most Responsive n-6 PUFA," *British Journal of Nutrition*, Vol. 103, No. 4, 2010, pp. 522-529. doi:10.1017/S0007114509992133

^{xiii} W. E. Lands, A. Morris and B. Libelt, "Quantitative Effects of Dietary Polyunsaturated Fats on the Composition of Fatty Acids in Rat Tissues," *Lipids*, Vol. 25, No. 9, 1990, pp. 505-516.

^{xiv} W. B. Zhang, P. B. Addis and T. P. Krick, "Quantification of 5 α -Cholestane-3 β , 5, 6 β -Triol and Other Cholesterol Oxidation Products in Fast-Food French-Fried Potatoes," *Journal of Food Science*, Vol. 56, No. 3, 1991, pp. 716-718. doi:10.1111/j.1365-2621.1991.tb05364.x

^{xv} W. Korytowski, G. J. Bachowski and A. W. Girotti, "Photoperoxidation of Cholesterol in Homogenous Solution, Isolated Membranes, and Cells: Comparisons of the 5 α - and 6 β -Hydroperoxides as Indicators of Singlet Oxygen Intermediacy," *Photochemistry and Photobiology*, Vol. 56, No. 1, 1992, pp. 1-8. doi:10.1111/j.1751-1097.1992.tb09594.x

^{xvi} G. Spiteller, "Is Atherosclerosis a Multifactorial Disease or Is It Induced by a Sequence of Lipid Peroxidation Reactions?," *Annals of the New York Academy of Sciences*, Vol. 1043, No. 1, 2005, pp. 355-366. doi:10.1196/annals.1333.042

^{xvii} G. Spiteller, "Peroxy Radicals: Inductors of Neurodegenerative and Other Inflammatory Diseases. Their Origin and How They Transform, Cholesterol, Phospholipids, Plasmalogens, Polyunsaturated Fatty Acids, Sugars, and Proteins into Deleterious Products," *Free Radical Biology and Medicine*, Vol. 41, No. 3, 2006, pp. 362-387. doi:10.1016/j.freeradbiomed.2006.03.013

^{xviii} Brenner, RR, "Hormonal modulation of Δ -6 and Δ -5 desaturases: case of diabetes," *Prostaglandins, Leukotrienes, and Essential Fatty Acids*, 68 (2003), 151-162; Mikhailidis, DP, et al., "The effect of dihomogammalinolenic on platelet aggregation and prostaglandin release, erythrocyte membrane fatty acids and serum lipids: evidence for defects in PGE1 synthesis and Δ -5-desaturase activity in insulin-dependent diabetics," *Diabetes Research* (1986) 3,7-12; Brown JE, Lindsay RM, Riemersma RA, "Linoleic acid metabolism in the spontaneously diabetic rat: Δ -6 desaturase activity vs. product / precursor ratios," *Lipids*. 2000 Dec;35(12):1319-23; Ray, TK, et al., "Regulation of insulin receptor activity of human erythrocyte membrane by prostaglandin E1 [PGE₁]," *Biochim Biophys Acta*. 1986 Apr. 25; 856(3):421-7; Mikhailidis, DP, et al., "The effect of dihomogammalinolenic on platelet aggregation and prostaglandin release, erythrocyte membrane fatty acids and serum lipids: evidence for defects in PGE1 synthesis and Δ -5 desaturase activity in insulin-dependent diabetics," *Diabetes Research* (1986) 3,7-12; Hissen, W, et al., Effect of prostaglandin E1 on platelet aggregation in vitro and in hemorrhagic shock," *Microvascular Research*, Vol 1, Issue 4, October 1969, pages 374-378; Weiss, C., et al., "Hemostasis and fibrinolysis in patients with intermittent claudication: effects of prostaglandin E1, Prostaglandins, Leukotrienes and Essential Fatty Acids, Nov. 2000; 63(5):271-277; Nakada, T, et al., "Membrane fatty acid composition shows a Δ -6 desaturase abnormality in Alzheimer's disease, *NeuroReport* 1, 153-155 (1990); Willard, DE, et al., "Identification of a fatty acid Δ -6 desaturase deficiency in human skin fibroblasts," *The Journal of Lipid Research*, 42, 2001, pages 501-508; Libby P. "Inflammation in atherosclerosis." *Nature*. 2002 Dec 19;420(6917):868-874; "A defect in the activity of Δ -6 and Δ -5 desaturases may be a factor in the initiation and progression of atherosclerosis," *Prostaglandins Leukot Essent Fatty Acids*. 2007;76(5):251-268; Savary, S, et al., "Fatty acids – Induced lipotoxicity and inflammation," *Current Drug Metabolism*, 2012, Vol. 13, No. 10, pages 1358-1370; Weiss, C, et al., "Hemostasis and fibrinolysis in patients with intermittent claudication: effects of prostaglandin E1," *Prostaglandins, Leukotrienes and Essential Fatty Acids*, Nov. 2000; 63(5):271-277; Lazaro, I, et al., "Linoleic Acid Status in Cell Membranes Inversely Relates to the Prevalence of Symptomatic Carotid Artery Disease," *Stroke*. 2021;52:703–

- 706; Ren, H-X, et al., Fang, W, et al., "Effect of prostaglandin E1 [PGE1] on TNF-induced vascular inflammation in human umbilical vein endothelial cells," *Can J Physiol Pharmacol*. 2010 May;88(5):576-83; Das, U, "A defect in the activities of Δ -6 and Δ -5 desaturase and pro-resolution bioactive lipids in the pathobiology of non-alcoholic fatty liver disease," *World Journal of Diabetes*, 2011 November 15:2(11).
- ^{xxix} Robert I Richards, Sarah A Robertson, Daniel L Kastner, "Neurodegenerative diseases have genetic hallmarks of autoinflammatory disease," *Human Molecular Genetics*, Volume 27, Issue R2, 01 August 2018, Pages R108–R118.
- ^{xx} Felton, CV, et al., "Dietary polyunsaturated fatty acids and compositions of human aortic plaque," *Lancet*.; 344:1195-1196, 1994.
- ^{xxi} Waddington E, et al., Identification and quantification of unique fatty acid and oxidative products in human atherosclerotic plaque using high-performance lipid chromatography," *Annals of Biochemistry*, 2001 May 15;292(2):234-44. doi: 10.1006/abio.2001.5075
- ^{xxii} Kühn H, et al., "Structure elucidation of oxygenated lipids in human atherosclerotic lesions," *Eicosanoids*, 1992;5(1):17-22.
- ^{xxiii} AHA Heartwire 2009, © 2009 Medscape, January 28, 2009 (Dallas, Texas), based on Journal of the American Heart Association. Ref.: AHA Science Advisory, Harris WS, et al., "Omega-6 fatty acids and risk for cardiovascular disease: a science advisory from the American Heart Association Nutrition Subcommittee of the Council on Nutrition, Physical Activity, and Metabolism; Council on Cardiovascular Nursing; and Council on Epidemiology and Prevention" downloaded from circ.ahajournals.org on January 29, 2009. Published in *Circulation*. 2009;119:902-907.
- ^{xxiv} Virtanen, JK, et al., "The associations of serum n-6 polyunsaturated fatty acids with serum C-reactive protein in men: the Kuopio Ischaemic Heart Disease Risk Factor Study," *European Journal of Clinical Nutrition*, online accessed November 18, 2017, <https://doi.org/10.1038/s41430-017-0>.
- ^{xxv} Bunting, S., Moncada, S., and Vane, J.R., "Prostaglandin–Thromboxane A2 Balance: Pathophysiological and Therapeutic Implications," *British Medical Journal*, (1993) Vol. 39, No. 3, pp 271-276.
- ^{xxvi} Weinberg RA, *One renegade cell: how cancer begins*, Basic Books, NY, NY 1998; Balkwill, F., et al.; "Smoldering and Polarized Inflammation in the Initiation and Promotion of Malignant Disease," *Cancer Cell*, Vol. 7, No. 3, pages 211-217, March 2005; "Distinct Role of Macrophages in Different Tumor Microenvironments," *Lewis, C. and Pollard, J., Cancer Research*, Vol. 66, No. 2, pages 605-612; January 15, 2006; "Paradoxical Roles of the Immune System during Cancer Development," *Visser, K., et al., Nature Reviews Cancer*, Vol. 6, No.1, pages 24-37; January 2006.
- ^{xxvii} Kiebish MA, Han X, Cheng H, Chuang JH, Seyfried TN. Cardiolipin and electron transport chain abnormalities in mouse brain tumor mitochondria: lipidomic evidence supporting the Warburg theory of cancer. *J Lipid Res* 2008;49:2545-66; Krebs JJ, Hauser H, Carafoli E. Asymmetric distribution of phospholipids in the inner membrane of beef heart mitochondria. *J Biol Chem* 1979;254:5308-16; Scottish Crop Research Institute (MRS Lipid Analysis Unit, Invergowrie, Dundee DD2 5DA, Scotland). Cardiolipin (diphosphatidylglycerol). Structure, occurrence, biology and analysis. Retrieved January 20, 2009, from: <http://www.lipidlibrary.co.uk/Lipids/dpg/index.htm>
- ^{xxviii} Sullivan, E. Madison, et al., "Docosahexaenoic acid lowers cardiac mitochondrial enzyme activity by replacing linoleic acid in the phospholipidome," *Journal of Biological Chemistry*, 2018, 293: 466-2018 Jan 12;293(2):466-483.
- ^{xxix} Halbleib, K., et al., "Activation of the Unfolded Protein Response by Lipid Bilayer Stress," *Molecular Cell*, Vol. 67, Issue 4, pp 673-684.e8, August 17, 2017.
- ^{xxx} I. M., D. N. Crozier and R. B. Caton, "Abnormal Fatty Acid Composition and Impaired Oxygen Supply in Cystic Fibrosis Patients," *Pediatrics*, Vol. 57, No. 4, 1976, pp. 480-486. doi:10.1054/plef.2000.0214
- ^{xxxi} G. Spiteller, "The Relation of Lipid Peroxidation Processes with Atherogenesis: A New Theory on Atherogenesis," *Molecular Nutrition & Food Research*, Vol. 49, No. 11, 2005, pp. 999-1013. doi:10.1002/mnfr.200500055
- ^{xxxii} U. N. Das, "A Defect in the Activity of D6 and D5 Desaturases May Be a Factor in the Initiation and Progression of Atherosclerosis," *Prostaglandins, Leukotrienes and Essential Fatty Acids*, Vol. 76, No. 5, 2007, pp.251-268. doi:10.1016/j.plefa.2007.03.001
- ^{xxxiii} Winkelstein, A and Kelly, VE, "The Pharmacologic Effects of PGE1 on Murine Lymphocytes," *Blood* (1980) 55 (3): 437-443.
- ^{xxxiv} C. Weiss, S. Regele, T. Velich, P. Bärtsch and T. Weiss, "Hemostasis and Fibrinolysis in Patients with Intermittent Claudication: Effects of Prostaglandin E1," *Prostaglandins Leukotrienes and Essential Fatty Acids*, Vol. 63, No. 5, 2000, pp. 271-277.
- ^{xxxv} F. Gao, H. Kim, M. Igarashi, et al., "Liver Conversion of Docosahexaenoic and Arachidonic Acids from Their 18-Carbon Precursors in Rats on a DHA-Free but α -LNA-Containing n-3 PUFA Adequate Diet," *Biochimica et Biophysica Acta*, Vol. 1811, No. 7-8, 2011 pp. 484-489. doi:10.1016/j.bbalip.2011.05.008
- ^{xxxvi} G. Barceló-Coblijn, E. J. Murphy, R. Othman, M. H. Moghadasian, T. Kashour and J. K. Friel, "Flaxseed Oil and Fish-Oil Capsule Consumption Alters Human Red Blood Cell n-3 Fatty Acid Composition: A Multiple-Dosing Trial Comparing 2 Sources of n-3 Fatty Acid," *The American Journal of Clinical Nutrition*, Vol. 88, No. 3, 2008, pp. 801-809. www.ncbi.nlm.nih.gov/pubmed/18779299
- ^{xxxvii} A. Welch, S. Shakya-Shrestha, M. Lentjes, N. J. Wareham and K. Khaw, "Dietary Intake and Status of N-3 Polyunsaturated Fatty Acids in a Population of Fish-Eating and Non-Fish-Eating Meat-Eaters, Vegetarians, and Vegans and the Precursor-Product Ratio of α -Linolenic Acid to Long-Chain N-3 Polyunsaturated Fatty Acids: Results From the EPIC-Norfolk [Cancer & Nutrition Study] Cohort," *The*

American Journal of Clinical Nutrition, Vol. 92, No. 5, 2010, pp. 1040-1051. doi:10.3945/ajcn.2010.29457

^{xxxviii} M. Plourde and S. C. Cunnane, "Extremely Limited Synthesis of Long-Chain Polyunsaturates in Adults: Implications for Their Dietary Essentiality and Use as Supplements," *Applied Physiology, Nutrition, and Metabolism*, Vol. 32, No. 4, 2007, pp. 619-634. doi:10.1139/H07-03

^{xxxix} C. V. Felton, D. Crook, M. J. Davies and M. F. Oliver, "Dietary Polyunsaturated Fatty Acids and Composition of Human Aortic Plaques," *The Lancet*, Vol. 344, No. 8931, 1994, pp. 1195-1196. doi:10.1016/S0140-6736(94)90511-8

^{xl} C. V. Felton, D. Crook, M. J. Davies and M. F. Oliver, "Relation of Plaque Lipid Composition and Morphology to the Stability of Human Aortic Plaques," *Arteriosclerosis, Thrombosis, and Vascular Biology*, Vol. 17, No. 7, 1997, pp. 1337-1345. doi:10.1161/01.ATV.17.7.1337

^{xli} V. Bertele, C. Cerletti and G. de Gaetano, "Pathophysiology of Critical Leg Ischaemia and Mode of Action of Prostaglandins," *Prostaglandins in the Cardiovascular System: Proceedings of the 5th International "Symposium on Prostaglandins in the Cardiovascular System"*, Vienna, 22-26 September 1991, pp. 18-26.

^{xlii} Willard, DE, et al., "Identification of a fatty acid Δ -6 desaturase deficiency in human skin fibroblasts," *The Journal of Lipid Research*, 42, 2001, pages 501-508; Chapkin RS, Ziboh VA, Marcelo CL, Voorhees JJ. "Metabolism of essential fatty acids by human epidermal enzyme preparations: evidence of chain elongation." *J Lipid Res* 1986; 27:945-954; Anderson A, Sjödin A, Hedman A, Olsson R, Vessby B. "Fatty acid profile of skeletal muscle phospholipids in trained and untrained young men." *Am J Physiol Endocrinol Metab* 2000;279:E744-E751; Albina E et al, "Detrimental effect of an ω -3 fatty-acid enriched diet on wound healing." *J Parenter Enteral Nutr*. 1993;17(6):519-521: "Current results show that substituting ω -3 fatty acid [fish oil] for ω -6 fatty acids in the diet is deleterious to the mechanical properties of wounds at 30 days."

^{xliii} Libby P. "Inflammation in atherosclerosis." *Nature*. 2002 Dec 19-26;420(6917):868-874.

^{xliv} Guo S, DiPietro LA. "Factors affecting wound healing." *J Dent Res*. 2010;89(3):219-229.

^{xlvi} Murray RK et al. *Harper's Illustrated Biochemistry*. 26th ed. New York: McGraw-Hill; 2003:97; Guyton AC, Hall JE. *Textbook of Medical Physiology*. 9th ed. W.B. Saunders Co.; 1996:16,861-862; Krebs, JJ, Hauser H, Carafoli E, "Asymmetric distribution of phospholipids in the inner membrane of beef heart mitochondria." *J Biol Chem*. 1979;254:5308-5316; Zhang M et al. "Gluing the respiratory chain together: cardiolipin is required for supercomplex formation in the inner mitochondrial membrane." *J Biol Chem*. 2002;277:43553-43556.

^{xlvii} Dines KC, et al., "Effectiveness of natural oils as sources of gamma-linolenic acid to correct peripheral nerve conduction velocity abnormalities in diabetic rats: modulation by thromboxane A2 inhibition." *Prostaglandins Leukot Essent Fatty Acids*. 1996 Sep;55(3):159-65.

^{xlviii} Asp ML et al. "Time-dependent effects of safflower oil [LA] to improve glycemia, inflammation and blood lipids in

obese, post-menopausal women with type 2 diabetes: A randomized, double-masked, crossover study." *Clin Nutr*. 2011 Aug;30(4):443-449.; Kahleova H et al. "Vegetarian diet-induced increase in linoleic acid [LA] in serum phospholipids is associated with improved insulin sensitivity in subjects with type 2 diabetes." *Nutr Diabetes* 2013;3(6)e75; Dutta-Roy A. "Effect of evening primrose oil feeding on erythrocyte membrane properties in diabetes mellitus." In: Horrobin D, ed. *Omega-6 Essential Fatty Acids: Pathophysiology and Roles in Clinical Medicine*. New York: Wiley-Liss; 1990:505-511; Ray TK, Dutta-Roy AK, Sinha AK, "Regulation of insulin receptor activity of human erythrocyte membrane by prostaglandin E1," *Biochim Biophys Acta*. 1986; 856(3):421-427.

^{xlviii} Das UN. "A defect in the activity of Δ -6 and D5 desaturases may be a factor in the initiation and progression of atherosclerosis." *Prostaglandins Leukot Essent Fatty Acids*. 2007;76(5):251-268; "[O]mega-6 PUFAs also have powerful anti-inflammatory properties that counteract any proinflammatory activity," say the advisory authors. 'It's incorrect to view the omega-6 fatty acids as "proinflammatory."' Ref.: Farvid MS et al. "Dietary linoleic acid [LA/ parent omega-6] and risk of coronary heart disease: a systematic review and meta-analysis of prospective cohort studies." *Circulation*. 2014;130:1568-1578; Terano T et al. "Effect of oral administration of highly purified eicosapentaenoic acid on platelet function, blood viscosity and red cell deformability in healthy human subjects." *Atherosclerosis*. 1983;46:321-331; Weiss, C., et al., "Hemostasis and fibrinolysis in patients with intermittent claudication: effects of prostaglandin E1, Prostaglandins, Leukotrienes and Essential Fatty Acids, Nov. 2000; 63(5):271-277; Lazaro, I, et al., "Linoleic Acid Status in Cell Membranes Inversely Relates to the Prevalence of Symptomatic Carotid Artery Disease," *Stroke*. 2021;52:703-706.

^{xlix} RS Mehta, et al., "High fish oil diet increases oxidative stress potential in mammary gland of spontaneously hypertensive rats," *Clin Exp Pharmacol Physiol* 1994 Nov;21(11):881-9.

^l S. Sethi, O. Ziouzenkova, H. Ni, D. D. Wagner, J. Plutzky and T. N. Mayadas, "Oxidized Omega-3 Fatty Acids in Fish Oil Inhibit Leukocyte-Endothelial Interactions through Activation of PPAR," *Blood*, Vol. 100, No. 4, 2002, pp. 1340-1356. doi:10.1182/blood-2002-01-0316

^{li} Tsuduki, K., et al., Long-term intake of fish oil increases oxidative stress and decreases lifespan in senescence-accelerated mice," *Nutrition* 27, (2011), pages 334-337.

^{lii} Hulbert, A.J., "Metabolism and Longevity: Is There a Role for Membrane Fatty Acids?" *Integrative and Comparative Biology*, Volume 50, Number 5, 808-817, 2010.

^{liii} S. G. Kaasgaard, S., G. Holmer, C.-E. Hoy, W. A. Behrens and J. L. Beare-Rogers, "Effects of Dietary Linseed Oil and Marine Oil on Lipid Peroxidation in Monkey Liver in Vivo and in Vitro," *Lipids*, Vol. 27, No. 10, 1992, pp. 740-745. doi:10.1007/BF02535843

^{liv} Calder, P., "Omega-3 Polyunsaturated Fatty Acids, Inflammation and Immunity," *Institute of Human Nutrition*,

University of Southampton, Bassett Crescent Campbell End, Southampton, UK. Presented at: The International Society for the Study of Fatty Acids and Lipids (ISSFAL) 4th Congress, which met on June 4-9, 2000, in Tsukuba, Japan.

^{lv} J. Delarue, F. Labarthe and R. Cohen, "Fish-Oil Supplementation Reduces Stimulation of Plasma Glucose Fluxes during Exercise in Untrained Males," *British Medical Journal of Nutrition*, Vol. 90, No. 4, 2003, pp. 777-786. doi:10.1079/BJN2003964

^{lvi} Spector, A.A., "Plasma Free Fatty Acids and Lipoproteins as Sources of Polyunsaturated Fatty Acid for the Brain," *Journal of Molecular Neuroscience*, Vol. 16, 2001, pages 159-165. "Most of the plasma free fatty acid (EFA) is derived from the triglycerides stored in the adipose tissue"; "Metabolism of essential fatty acids by human epidermal enzyme preparations: evidence of chain elongation," R.S. Chapkin, et. al., *Journal of Lipid Research*, Volume 27, pages 954-959,

1986, and Agneta Anderson, et. al., *American Journal of Endocrinological Metabolism*, 279: E744-E751.

^{lvii} J. C. Umhau, W. Zhou, R. E. Carson, et al., "Imaging Incorporation of Circulating Docosahexaenoic Acid [DHA] into the Human Brain Using Positron Emission Tomography," *Journal of Lipid Research*, Vol. 50, No. 7, 2009, pp. 1259-1268. doi:10.1194/jlr.M800530-JLR200

^{lviii} P. L. Goyens, M. E. Spilker, P. L. Zock, M. B. Katan, and R. P. Mensink, "Conversion of Alpha-Linolenic Acid in Humans Is Influenced by the Absolute Amounts of Alpha-Linolenic Acid and Linoleic Acid in the Diet and Not by Their Ratio," *The American Journal of Clinical Nutrition*, 2006, Vol. 84, No. 1, pp. 44-53.

^{lix} N. Hussein, E. Ah-Sing, P. Wilkinson, C. Leach, B. Griffin and D. Millward, "Physiological Compartmental Analysis of Alpha-Linolenic Acid Metabolism in Adult Humans," *Journal of Lipid Research*, Vol. 46, 2005, pp. 269-280. doi:10.1194/jlr.M400225-JLR200